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CHAPTER 2

GENIC MOLECULAR MARKERS IN PLANTS: DEVELOPMENT AND APPLICATIONS

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Abstract: The current advancement in plant biology research encompassing: generation of huge amount of molecular-genetic data, development of impressive methodological skills in molecular biology experimentation, and systems analyses, has set the stage to search for ways/means to utilize the available resources to strengthen interdisciplinary efforts to find solutions to the challenging goals of plant breeding efforts (such as abiotic stress tolerance) ultimately leading to gainful applications in crop improvement. A positive fall out of such a realization and efforts has been the identification/development of a new class of very useful DNA markers called genic molecular markers (GMMs) utilizing the ever-increasing archives of gene sequence information being accumulated under the EST sequencing projects on a large number of plant species in the recent years. These markers being part of the cDNA/EST-sequences, are expected to represent the functional component of the genome i.e., gene(s), in contrast to all other random DNA-based markers (RDMs) that are developed/generated from the anonymous genomic DNA sequences/domains irrespective of their genic content/information. Therefore, identifying DNA sequences that demonstrate large effects on adaptive plant behavior remains fundamental to the development of GMMs. The few recent studies have now demonstrated the utility of these markers in genetic studies, and also shown that GMMs may be superior than RDMs for use in the marker-assisted selection, comparative mapping, and exploration of the functional genetic diversity in the germplasm adapted to different environments. The only constraint of GMMs is their low level of polymorphism as compared to the RDMs, which is expected of their origin from the relatively conserved functional portion of the genome. This chapter provides a critical review of the development and various applications of the GMMs.

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1. MOLECULAR MARKERS IN PLANT BREEDING

In agriculture, one of the main objectives of plant breeder is to improve the existing cultivars, which are deficient in one or more traits by crossing such cultivars with lines that possess the desired trait. A conventional breeding programme thus involves crossing whole genomes followed by selection of the superior recombinants from among the several segregation products. Indeed, such a procedure is laborious and time consuming, involving several crosses, several generations, and careful phenotypic selection, and the linkage drag (tight linkage of the undesired loci with the desired loci) may make it further difficult to achieve the desired objective. Advent of DNA marker technology, development of several types of molecular markers and molecular breeding strategies offered possibilities to plant breeders and geneticists to overcome many of the problems faced during conventional breeding.

Molecular markers are now widely used to track loci and genome regions in several crop-breeding programmes, as molecular markers tightly linked with a large number of agronomic and disease resistance traits are available in major crop species (Phillips and Vasil 2001, Jain et al. 2002, Gupta and Varshney 2004). These molecular markers include: (i) hybridization-based markers such as restriction fragment length polymorphism (RFLP), (ii) PCR-based markers: random amplification of polymorphic DNA (RAPD), amplified fragment length polymorphism (AFLP) and microsatellite or simple sequence repeat (SSR), and (iii) sequence-based markers: single nucleotide polymorphism (SNP). The majority of these molecular markers has been developed either from genomic DNA libraries (e.g. RFLPs and SSRs) or from random PCR amplification of genomic DNA (e.g. RAPDs) or both (e.g. AFLPs). These DNA markers can be generated in large numbers and can prove to be very useful for a variety of purposes relevant to crop improvement. For instance, these markers have been utilized extensively for the preparation of saturated molecular maps (genetical and physical). Their association with genes/QTLs controlling the traits of economic importance has also been utilized in some cases for indirect marker-assisted selection (MAS) (e.g. Koebner 2004, Korzun 2002). Other uses of molecular markers include gene introgression through backcrossing, germplasm characterization, genetic diagnostics, characterization of transformants, study of genome organization and phylogenetic analysis (see Jain et al. 2002). For plant breeding applications, SSR markers, among different classes of the existing markers, have been proven and recommended as markers of choice (Gupta and Varshney 2000). RFLP is not readily adapted to high sample throughput and RAPD assays are not sufficiently reproducible or transferable between laboratories. While both SSRs and AFLPs are efficient in identifying polymorphisms, SSRs are more readily automated (Shariflou et al. 2001). Although AFLPs can in principle be converted into simple PCR assays (e.g. STSs), this conversion can become cumbersome and complicated as individual bands are often composed of multiple fragments (Shan et al. 1999), particularly in large genome templates.

01 **2. GENIC MOLECULAR MARKERS: INTRODUCTION**
02 **AND DEVELOPMENTS**

03 Due to emphasis on functional genomics, several gene discovery projects in the
04 form of genome sequencing, transcriptome sequencing or gene expression studies
05 have been established since last five years. As a result, a large number of genes have
06 been identified through 'wet lab' as well as *in silico* studies and a wealth of sequence
07 data have been accumulated in public databases (e.g. <http://www.ncbi.nlm.nih.gov>;
08 <http://www.ebi.ac.uk>) in the form of BAC (bacterial artificial chromosome) clones,
09 ESTs (expressed sequence tags), full length cDNA clones and genes. The availability
10 of enormous amount of sequence data from complete or partial genes has made it
11 possible to develop the molecular markers directly from the parts of genes. These
12 markers are referred as "genic" molecular markers (GMM).

13 The majority of the markers, developed and used in the past as described above
14 in section 1, are directly derived from the genomic DNA, and therefore could
15 belong to either the transcribed or the non-transcribed part of the genome without
16 any information available on their functions. In contrast, GMMs developed from
17 coding sequences like ESTs or fully characterized genes frequently have been
18 assigned known functions. Based on the site of polymorphism and later's effect on
19 phenotypic variation, GMMs have been classified into two groups (Anderson and
20 Luebberstedt 2003):

- 21 (i) Gene-targeted markers (GTMs): derived from polymorphisms within genes,
22 however not necessarily involved in phenotypic trait variation, e.g. untranslated
23 regions (UTRs) of EST sequences (Schmitt et al. 2006; Aggarwal et al 2007);
24 (ii) Functional markers (FMs): derived from polymorphic sequences or sites within
25 genes and, thus, more likely to be causally involved in phenotypic trait
26 variation (e.g. candidate gene-based molecular markers). The FMs, depending
27 on the involvement in the phenotypic trait variation, are further classified
28 into two subgroups: (a) indirect functional markers (IFMs), for which the role
29 for phenotypic trait variation is indirectly known, and (b) direct functional
30 markers (DFMs), for which the role for the phenotypic trait variation is well
31 proven.

32 As per the above terminology, the molecular markers derived from anonymous
33 regions of the genome are called random DNA markers (RDMs), which may or
34 may not be developed from the polymorphic site in gene or may not be developed
35 from a gene at all.

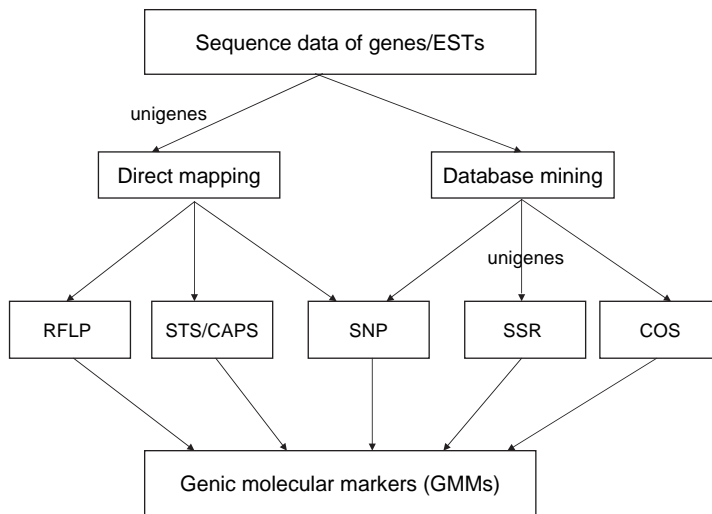
36 Although genic markers were developed earlier also, these were in the form
37 of cDNA-RFLP (Graner et al. 1991, Causse et al. 1994) for which functions
38 could not be predicted at that time. However, some efforts were made to sequence
39 these early cDNA clones to determine the genes and their functions (Michalek
40 et al. 1999). Compared to these earlier efforts, development of genic markers have
41 become a reality only in recent years, because of accumulation of large ESTs or
42 gene sequences resources resulting from EST and genome sequencing projects in
43 several crop species and also due to the developments in the field of bioinforma-
44 tics (Gupta and Rustgi 2004). For example, several transcriptome resources have

01 become available (http://www.ncbi.nlm.nih.gov/dbEST/dbEST_summary.html),
 02 and software tools or perl scripts have been developed to search for SSRs and
 03 SNPs from EST or gene sequences (Varshney et al. 2004, 2005a).

04 Although, whole genome sequencing and annotation is the way to identify the
 05 entire gene repository of a species, this has been possible only for a limited number
 06 of crop species involving large scale sequencing of their genome or gene space. On
 07 the other hand, ESTs represent a basic commodity within the analysis of genomes
 08 and their genes for a species (Rudd et al. 2003). Whereas the complete sequencing
 09 of a genome may utilize either a clone-by-clone approach or a whole genome
 10 shotgun approach to acquire adequate coverage to assemble a meaningful scaffold,
 11 EST sequencing is directed at the quick, cheap and simple sequencing of partial
 12 gene transcripts (Sreenivasulu et al. 2002). As a result, a significant redundancy
 13 can be observed in gene sequence data obtained from EST sequencing projects (see
 14 Varshney et al. 2004). Therefore before developing molecular markers from ESTs,
 15 it is essential to define the “unigenes” after cluster analysis of random ESTs using
 16 appropriate computer programmes such as stackPack (Miller et al. 1999).

17 Once the unigene sequence data from EST analysis or non-redundant set of genes
 18 are available, molecular markers can be developed using two main approaches:

19 (1) Direct mapping: Under this approach, either the cDNA clones corresponding
 20 to the ESTs of interest can be used as RFLP probe or the PCR primers can be
 21



39 *Figure 1.* A scheme for development of genic molecular markers (GMMs). Two common ways to
 40 develop GMMs are shown in the figure. In the first method, the sequence data are used to define the
 41 unigenes and then the cDNA clones or genic clones corresponding to the unigenes can be assayed
 42 as RFLPs or the unigene sequence data can be used to design the primer pairs and assayed using
 43 STS/CAPS or SNP assays. In the second method, the sequence data can be mined by using some
 44 computer programmes or scripts to identify the SSRs, SNPs or COSs from given sequence data and
 then these markers, after defining the unigenes, can be assayed using appropriate genotyping platforms

01 designed for the EST/gene and used as STS or CAPS marker. Direct mapping
02 approach should be undertaken with the unigene set of ESTs or genes only.

03 (2) *In silico* mining: In this approach, the SSR or SNP identification software
04 tools are used to screen the sequence data for ESTs/genes. For identification of
05 SNPs, the redundant set of EST data, generated from more than one genotype
06 of a given species, are used. However, after identification of SNPs, only non-
07 redundant set of ESTs should be considered for SNP mapping.

08 A scheme for development of GMMs has been shown in Figure 1. Development of
09 FMs, however, requires: (i) functionally characterized genes, (ii) allele sequences
10 from such genes, (iii) identification of polymorphic, functional motifs affecting
11 plant phenotype within these genes, and (iv) validation of associations between
12 DNA polymorphisms and trait variation. Therefore depending on the objective as
13 well as available information or feasibility, the FMs, the special class of GMMs,
14 can also be generated.

16 3. APPLICATIONS OF GENIC MOLECULAR MARKERS

17
18 Molecular markers have already shown their applications in a variety of ways in
19 several plant species (see Gupta and Varshney 2004). The development of GMMs,
20 now permits a targeted approach for detection of nucleotide diversity in genes
21 controlling agronomic traits in plant populations. Some main areas of plant breeding
22 and genetics, where the implementation of GMMs will prove quite useful, are
23 discussed here.

25 3.1. Trait Mapping

26
27 One of the main applications of molecular markers in plant breeding is their use as
28 diagnostic markers for the trait in the selection. However, use of random molecular
29 markers (RDMs) as a diagnostic tool entails the risk of losing the linkage through
30 genetic recombination. Even in case of GMMs, the gene-targeted markers (GTMs)
31 where polymorphism was discovered through one allele analysis without any further
32 specification of the polymorphic sequence motif are threatened by the same way
33 (Rafalski and Tingey, 1993). In contrast to RDMs or GTMs, FMs (DFMs or IFMs)
34 allow reliable application of markers in populations without prior mapping and the
35 use of markers in mapped populations without risk of information loss owing to
36 recombination.

37 The development of FMs is expensive and cannot be undertaken for all the traits
38 and in all crop species, GMM have been developed and mapped in several plant
39 species (Table 1). The genetic maps, developed after mapping/integration of GMM
40 are called “transcript” or “gene” maps. For example, based on the candidate genes
41 for drought tolerance, a comprehensive set of >200 gene-based markers have been
42 developed for barley (Rostocks et al. 2005). Recently, a “transcript map” of barley
43 after integrating more than 1000 gene-based markers (GTMs) has been developed,
44 (Stein et al. 2007). A kind of transcriptome map based on deletion mapping of

01 *Table 1.* Some reports on development of genic molecular markers in important plant species

02 General name	03 Species	04 Type of markers developed	05 References
06 Cereals and grasses			
07 Barley	<i>Hordeum vulgare</i>	EST-SSR, EST-SNP, EST-RFLP, cDNA-RFLP	Thiel et al. 2003, Rostocks et al. 2005, Varshney et al. 2006, Willsmore et al. 2006, Stein et al. 2007, Varshney et al. 2007b
10 Maize	<i>Zea mays</i>	cDNA-RFLP, EST-SNP	Gardiner et al. 1993, Chao et al. 1994, Picoult-Newberg et al. 1999, Falque et al. 2005
12 Wheat	<i>Triticum aestivum</i>	EST-SSR, EST-SNP, cDNA-RFLP	Holton et al. 2002, Yu et al. 2004, Somers et al. 2003, Gao et al. 2004, Qi X. et al. 2004, Nicot et al. 2004
16 Rice	<i>Oriza sativa</i>	EST-SSR, EST-SNP, cDNA-RFLP, Intron Length Polymorphism (ILP)	Causse et al. 1994, Harushima et al. 1998, Temnykh et al. 2001, Feltus et al. 2004, Wang et al. 2005
19 Rye	<i>Secale cereale</i>	EST-SSR, EST-SNP	Hackauf and Wehling, 2002, Khlestkina et al. 2004, Varshney et al. 2007b
22 Sorghum	<i>Sorghum bicolor</i>	EST-SSR, cDNA-RFLP	Childs et al. 2001, Klein et al. 2003, Bowers et al. 2003, Ramu et al. 2006, Jayashree et al. 2006
25 Lolium	<i>Lolium perenne</i>	EST-SSR	Faville et al. 2004
26 Legumes			
27 White clover	<i>Trifolium repens</i>	EST-SSR	Barret et al. 2004
28 Soybean	<i>Glycine max</i>	EST-SSR	Song et al. 2004, Zhang et al. 2004
29 Fiber and oil seed crops			
31 Cotton	<i>Gossypium</i> sps.	EST-SSR	Zhang et al. 2005, Chee et al. 2004, Park et al. 2005- Lai et al. 2005
32 Sunflower	<i>Helianthus</i> sps.	EST-SNP	Lai et al. 2005
33 Fruit and vegetables			
34 Grape	<i>Vitis vinifera</i>	EST-SSR	Chen et al. 2006
35 Kiwi fruit	<i>Actinidia chinensis</i>	EST-SSR	Fraser et al. 2004
36 Raspberry	<i>Rubus</i> spp.	EST-SSR	Graham et al. 2004
37 Tomato	<i>Lycopersicon esculentum</i>	EST-SSR	Frary et al. 2005
38 Strawberry	<i>Fragaria</i> spp.	EST-SSR	Sargent et al. 2006
39 Trees			
41 Pinus	<i>Pinus</i> spp.	EST-SSR, ESTP	Cato et al. 2001
42 Coffee	<i>Coffea</i> spp.	EST-SSR	Bhat et al. 2005, Aggarwal et al. 2007

01 more than 16,000 gene loci has been developed in wheat (Qi L-L et al. 2004). Such
02 molecular maps, not only provide gene based molecular markers associated with
03 the trait of interest after the QTL analysis, but also can be compared with those of
04 the other related plant species in an efficient manner.

06 **3.2. Functional Diversity**

08 Characterization of genetic variation within natural populations and among breeding
09 lines is crucial for effective conservation and exploitation of genetic resources
10 for crop improvement programmes. Molecular markers have proven useful for
11 assessment of genetic variation in germplasm collections (Hausmann et al. 2004;
12 Maccaferri et al. 2006). Evaluation of germplasm with GMMs might enhance the
13 role of genetic markers by assaying the variation in transcribed and known function
14 genes, although there may be a higher probability of bias owing to selection.

15 While using the genic SSR markers for diversity studies, the expansion and
16 contraction of SSR repeats in genes of known function can be tested for association
17 with phenotypic variation or, more desirably, biological function (Ayers et al. 1997).
18 The presence of SSRs in the transcripts of genes suggests that they might have a role in
19 gene expression or function; however, it is yet to be determined whether any unusual
20 phenotypic variation might be associated with the length of SSRs in coding regions as
21 was reported for several diseases in human (Cummings and Zoghbi 2000). Similarly,
22 the use of SNP markers for diversity studies may correlate the SNPs of coding *vs.* non-
23 coding regions of the gene with the trait variation. The variation associated with deleterious
24 characters, however, is less likely to be represented in the germplasm collections
25 of crop species than among natural populations because undesirable mutations are
26 commonly culled from breeding populations (Cho et al. 2000).

27 Several studies involving GMMs, especially genic SSRs, have been found useful
28 for estimating genetic relationship on one hand (see Gupta et al. 2003 Gupta and
29 Rustgi 2004, Varshney et al. 2005a) while at the same time these have provided
30 opportunities to examine functional diversity in relation to adaptive variation (Eujayl
31 et al. 2001, Russell et al. 2004). It seems likely that with the development of
32 more GMMs in major crop species, genetic diversity studies will become more
33 meaningful by a shift in emphasis from the evaluation of anonymous diversity to
34 functional genetic diversity in the near future. Nevertheless, use of the neutral RDM
35 markers will remain useful in situations where: (i) GMMs would not be available,
36 and (ii) to address some specific objectives e.g. neutral grouping of germplasm.

38 **3.3. Interspecific or Intergeneric Transferability**

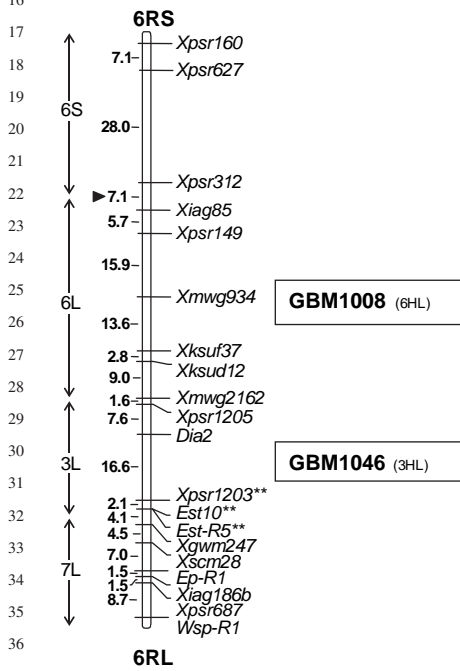
40 Perhaps one of the most important features of the GMMs is that these markers
41 provide high degree of transferability among distantly related species. In contrast,
42 except RFLPs all other RDMs are generally constrained in this regard. Transferability
43 of GMM markers to related species or genera has now been demonstrated
44 in several studies (Table 2). For example, a computational study based on analysis

01 Table 2. Some examples of interspecific or intergeneric transferability of genic molecular markers

02 Plant species	03 Marker type	04 Species, recorded transferability	05 Reference
Cereals and grasses			
06 Barley (<i>Hordeum vulgare</i>)	EST-SSR, EST-SNP	Wheat, rice, rye	Thiel et al. 2003, Varshney et al. 2004, 2007b
07 Wheat (<i>Triticum aestivum</i>)	EST-SSR	08 <i>Aegilops</i> and <i>Triticum</i> species, barley, maize, rice, rye, oats, soybean, <i>Lophopyrum elongatum</i>	Holton et al. 2002, Gupta et al. 2003, Gao et al. 2003, Bandopadhyay et al. 2004, Yu et al. 2004, Mullan et al. 2005, Tang et al. 2006
11 Rice (<i>Oryza sativa</i>)	EST-SSR	wild species of rice	Cho et al. 2000
12 Sugarcane (<i>Saccharum officinarum</i>)	EST-SSR	<i>Saccharum robustum</i> , <i>Erianthus</i> and Sorghum	Cordeiro et al. 2001
15 Sorghum (<i>Sorghum bicolor</i>)	EST-SSR	<i>Eleusine coracana</i> , <i>Seashore paspalum</i> , finger millet	Wang et al. 2005
16 Tall fescue (<i>Festuca</i>)	EST-SSR	subfamilies of Poaceae	Mian et al. 2005
Fiber and oilseed crops			
18 Cotton (<i>Gossypium hirsutum</i>)	EST-SSR	Cotton species	Saha et al. 2003
20 Sunflower (<i>Helianthus annuus</i>)	EST-SSR	<i>Heliantus angustifolius</i> , <i>Helianthus verticillatus</i>	Pashley et al. 2006
Fruit and vegetables			
24 Strawberry (<i>Fragaria vesca</i>)	EST-SSR	<i>F. gracilis</i> , <i>F. iinumae</i> , <i>F. nilgerrensis</i> , <i>F. nipponica</i>	Bassil et al. 2006
26 Apricot (<i>Prunus armeniaca</i>)	EST-SSR	Vitaceae and Roseaceae family	Decroocq et al. 2003
27 Grape (<i>Vitis vinifera</i>)	EST-SSR	> 25 species from 5 Vitaceae and Roseaceae	Scott et al. 2000, Rossetto et al. 2002, Arnold et al. 2002, Decroocq et al. 2003
30 Tomato (<i>Solanum lycopersicum</i>)	EST-SSR	Solanaceous members	Frary et al. 2005
Ferns and trees			
33 Alpine lady-fern (<i>Athyrium distentifolium</i>)	EST-SSR	9 species from Woodsiaceae	Woodhead et al. 2003
35 Pinus (<i>Pinus taeda</i>)	EST-SSR	12 <i>Pinus</i> species	Komulainen et al. 2003, Changne et al. 2004, Liewlaksaneeyanawin et al. 2004
38 Spruce (<i>Picea glauca</i>)	EST-SSR	23 <i>Picea</i> species	Rungis et al. 2004
40 Citrus (<i>Citrus sinensis</i>)	EST-SSR	<i>Poncirus trifoliata</i>	Chen et al. 2006
42 Coffee (<i>Coffea arabica</i> , <i>Coffea canephora</i>)	EST-SSR	16 species of coffee and <i>Psilanthus</i>	Bhat et al. 2005, Poncet et al. 2006, Aggarwal et al. 2007

01 of ~1000 barley GMMs suggested a theoretical transferability of barley markers
 02 to wheat (95.2%), rice (70.3%), maize (69.3%), sorghum (65.9%), rye (38.1%) and
 03 even to dicot species (~16%). Infact, *in silico* analyses of GMMs of wheat, maize
 04 and sorghum with complete rice genome sequence data have provided a larger
 05 number of anchoring points among different cereal genomes as well as provided
 06 insights into cereal genome evolution (Sorrells et al. 2003, Salse et al. 2004).

07 In some studies, the use of GMMs of major crop species has been shown to enrich the
 08 genetic maps of related plant species for which little marker information is available.
 09 For example, barley EST-SSR as well as EST-SNP markers have been shown trans-
 10 ferable as well as mappable in syntenic regions of rye (Varshney et al. 2004, 2005c,
 11 2007a; Figure 2). Further, such kind of markers from the related plant species offers the
 12 possibility to develop anchor or conserved orthologous sets (COS) for genetic analysis
 13 and breeding in different species. In this direction, Rudd et al. (2005) identified a large
 14 repository of such COS markers and developed a database called "PlantMarker".



38 *Figure 2.* An example of integration of barley genic (EST-SSR) markers into syntenic regions of rye
 39 genetic map. Integrated barley markers (GBM1008, GBM1046) are shown in bold and capital font in
 40 boxes on right hand side. Details about other markers present on this linkage group are available in
 41 Korzun et al. (2001). Genetic distances are given in centimorgans (cM) on left hand side. The black
 42 triangle indicates the estimated centromere position. The relationship of the linkage group 6R in terms
 43 of Triticeae linkage group is shown on very left hand side (left to black triangle) as per Devos et al.
 44 (1993). Both barley genic markers from linkage group 3H and 6H are mapped into expected syntenic
 regions of the rye linkage group 6R. S = short arm, L = long arm

01 4. COMPARISON OF GMMs AND RDMs

02 Since the development of first molecular markers i.e. RFLPs in 1980 (Botstein
03 et al. 1980), a diverse array of molecular marker technologies have come into
04 being revolutionizing conventional plant breeding efforts for crop improvement.
05 Significant strides have been made in crop improvement through conventional
06 random molecular markers (RDMs). For instance, these molecular markers besides
07 throwing light on organization, conservation and evolution of plant genomes, have
08 also aided geneticists and plant breeders to tag genes, map QTLs for the traits
09 of economic importance. Still, most of them are “anonymous” markers, that is to
10 say their biological function is unknown. In comparison, a putative function for
11 majority of the molecular markers, derived from the genes or ESTs, however can
12 be deduced using some bioinformatics tools; such markers (GMMs) are commonly
13 referred as functional markers (Varshney et al. 2005b). Although, in *stricto* sense,
14 the functional markers are based on functionally defined genes underlying specific
15 biochemical or physiological functions and therefore the FMs can be considered as
16 a class of GMMs (Anderson and Lueberstedt 2003).

17 The GMMs, like RDMs, could detect both length and sequence polymorphisms
18 in expressed regions of the genome but provide relatively stronger and robust
19 marker assays. However, as compared to the RDMs the developmental costs of
20 GMMs, depend on which specific class of GMMs is to be developed. Similarly
21 the applied value of the GMMs as compared to the RDMs varies depending on
22 the class of the GMMs. These relative costs and applications issues have been
23 detailed in Table 3. In summary, if the GMMs based on the polymorphic site
24 and verification are developed (i.e. FMs), these markers are superior to RDMs
25 for using them as diagnostic tools in marker-assisted selection as they may owe
26 the complete linkage with the trait locus alleles (Anderson and Lueberstedt
27 2003). In plant breeding, the GMMs are superior to RDMs for selection of, e.g.,
28 parent materials to build segregating populations, as well as subsequent selection
29 of lines (line breeding) or inbreds (hybrid breeding). Depending on the mode
30 of the GMM characterization, these can also be applied to the targeted combi-
31 nation of alleles in hybrid and synthetic breeding. In population breeding and
32 recurrent selection programs, the GMMs can be employed to avoid genetic drift at
33 characterized loci.

34 Being originated from the conserved proportion of the genome, the GMMs, as
35 compared to the RDMs, are the candidate markers for interspecific/intergeneric
36 transferability and comparative mapping/genomics studies in related plant species.
37 Since the GMMs represent the expressed portion of the genome, they sample the
38 variation in transcribed regions of the genome, and provide a more direct estimate
39 of functional diversity while screening the markers on the germplasm adapted
40 to different environments. Nevertheless, the GMMs, as compared to the RDMs
41 are less polymorphic and provide less alleles and lower PIC values. Additionally,
42 due to biased distribution in the genome, the GMMs are unsuitable for analyzing
43 population structure.
44

01 *Table 3.* Comparison of genic molecular markers (GMMs) with random DNA markers (RDMs)

02 Feature	GMMs	RDMs		
		03 gSSRs, SNPs	RFLPs	04 RAPD/AFLP/ ISSR etc.
05 Need for sequence data	06 Genes/ESTs data Essential	Essential	Not required	Not required
07 Costs of generation	Low*	High	High	Low-moderate
08 Labour involved	Less	Much	Much	Less
09 Level of polymorphism	Low	High	Low	Low-moderate
10 Interspecific 11 transferability and 12 comparative 13 mapping	High	Low-moderate	Moderate-High	Low-moderate
14 Function of markers	Known majority of times	Unknown majority of times	Unknown	Unknown
15 Utility in 16 marker- 17 assisted 18 selection	Great, if the marker is derived from the gene, involved in 19 expression of 20 trait	High	Moderate	Low-moderate

21
22 *generally GMMs are by products of the available transcriptome resources being developed for functional
23 genomic studies.

24

25 5. FUTURE DIRECTIONS OF GENIC MOLECULAR MARKERS

26

27 It is clear that the GMMs and especially the FMs are extremely useful source of
28 markers in plant breeding for marker-assisted selection because these markers may
29 represent the genes responsible for expression of target traits. If so, there will not
30 be any recombination between the markers and the trait, thus representing perfect
31 indirect selection tools. While low level of polymorphism is an inherent feature of
32 the GMMs, it is compensated by their higher interspecific transferability as well as
33 capacity to sample the functional diversity in the germplasm. These features make
34 the development and application of the GMMs more attractive for plant breeding
35 and genetics.

36 With more DNA sequence data being generated continuously, the trend is
37 towards cross-referencing genes and genomes using sequence and map-based tools.
38 Because polymorphism is a major limitation for many species, SSR- and SNP-
39 based GMMs will be valuable tools for plant geneticists and breeders. In the
40 longer term, development of allele-specific, functional markers (FMs) for the
41 genes controlling agronomic traits will be important for advancing the science
42 of plant breeding. In this context genic SSR and SNP markers together with
43 other types of markers that target functional polymorphisms within genes will be
44 developed in near future for major crop species. The choice of the most appropriate

01 marker system, however, needs to be decided on a case-by-case basis and will
 02 depend on many issues including the availability of technology platforms, costs
 03 for marker development, species transferability, information content and ease of
 04 documentation.

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